High-Resolution Cryo-Electron Tomography – Current State and Future Prospects

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Why Structural Biology?

- Structure and function of macromolecules are highly interdependent.
- Multiple structures of different functional states help to understand molecular mechanisms.
- *In situ* structures help to understand how molecules interact with their native environment.
- Structures can be used as starting point to design functional experiments (e.g. by guided design of mutations).

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Common Methods in Structural Biology

X-Ray Crystallography

- Can be very quick and easy.
- No principle limitations for protein size etc.
- Large amounts of highly pure material necessary.
- Protein needs to form highly ordered crystals.
- Phase problem needs to be solved.

NMR Spectroscopy

- True solution method.
- Accounts for dynamics in macromolecules.
- In-cell NMR is possible in some cases.
- Requires labelling with heavy nuclides (N2, C13, N15).
- Limited to small proteins or selective labelling.
- Sample needs to be extraordinarily stable.

Cryo Electron Microscopy (Single Particle Analysis)

- Lower sample amounts necessary.
- Proteins do not need to be crystallized.
- Not always easy to find good conditions for sample preparation.
- Sample preparation might be harmful for the protein.
- Difficult for smaller molecules.



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Artefacts



North O Providence O Providence

1500 tomograms with thickness below 150 nm.

Current performance

Approx. 10 lamellae per Day (8h)

Approx. one month for one dataset

Approx. two month of data acquisition

Current limitations:

- 1. Reliable vitrification
- 2. Artefact free transfer.
- 3. Reduced contamination rate.
- 4. Automated milling missing.
- 5. Faster data acquisition.

Target performance

30 lamellae per day

Approx. 10 days for one dataset

Approx. two weeks of data acquisition

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[1] A. Rigort (2016). Recent developments in FEI's in situ cryo-electron tomography workflow. *Nature Methods*. 13, 958-960.

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